

**C-REACTIVE PROTEIN (CRP)**  
**(Human)**  
**ELISA Kit Protocol**

(Cat. No.:EK-072-62)



**PHOENIX PHARMACEUTICALS, INC.**



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**CAUTION:**

Investigational device. Limited by law to investigational use. For research use only. Not for use in diagnostic procedures.

## INTRODUCTION AND PROTOCOL OVERVIEW

Human C-Reactive Protein (CRP) is an acute phase protein in humans and rabbits that has the ability to bind to a number of biologically important ligands such as histones and polycations. Expression of C-Reactive Protein is regulated mainly at the transcription level of Interleukin-6. The main biological function of C-Reactive Protein is to be host defense against bacterial pathogens and clearance of apoptotic and necrotic cells. In addition, C-Reactive Protein also participates in atherogenesis and pathogenesis of myocardial infarction.

Phoenix Pharmaceutical's Human C-Reactive Protein ELISA Kit is designed to measure the concentration of Human C-Reactive Protein from Human serum/plasma, or conditioned medium.

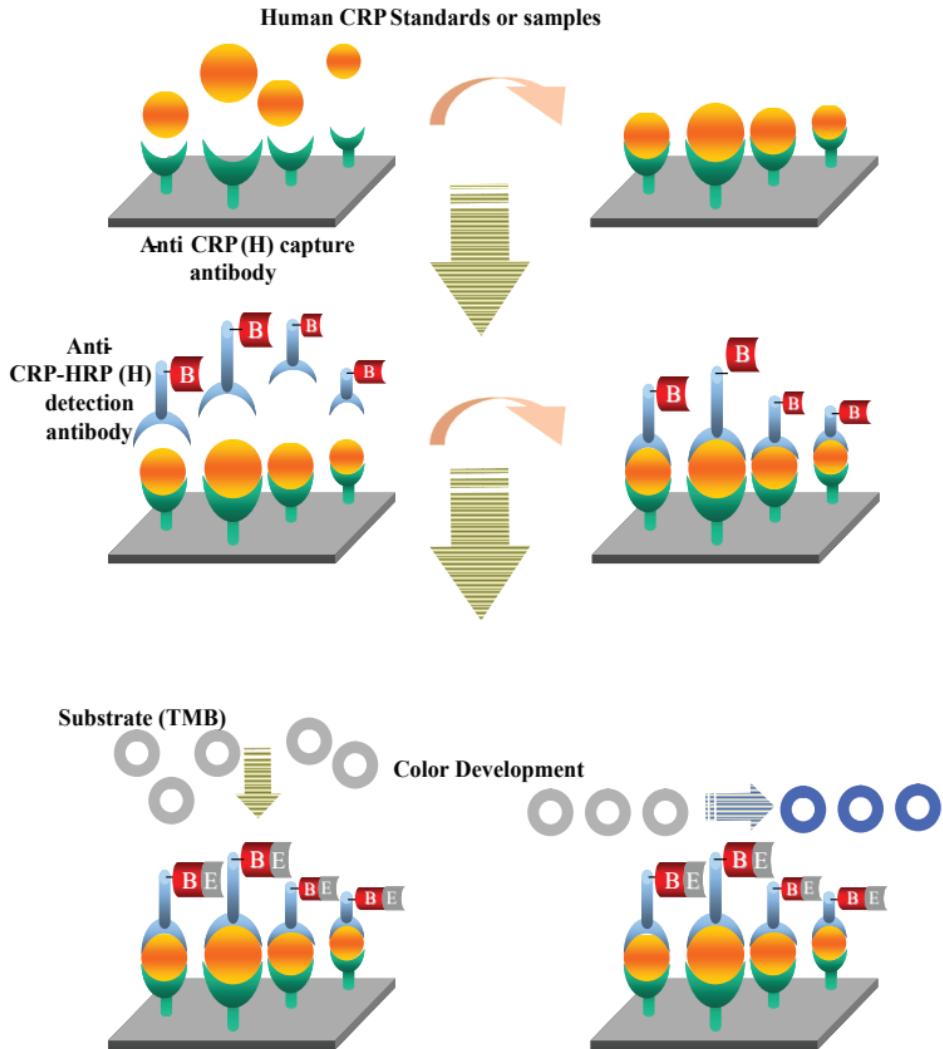
The immunoplate in this kit is pre-coated with Anti-Human C-Reactive Protein Capture Antibody and the nonspecific binding sites are blocked. The Human C-Reactive Protein in the sample or in the standard solution can bind to the capture antibody immobilized in the wells. After washing procedure, the biotinylated anti-Human C-Reactive Protein streptavidin-Horseradish Peroxidase Detection Antibody which can bind to the Human C-Reaction Protein trapped in the wells is added. After washing, the Streptavidin-Horseradish Peroxidase (SA-HRP) catalyzes the substrate solution (TMB). The exzyme-substrate reaction is terminated by the addition of a stop solution. The intensity of the color is directly proportional to the amount of Human C-Reactive Protein in the standard solutions or samples. A standard curve of Human C-Reactive Protein with known concentration can be established accordingly. The Human C-Reactive Protein with unknown concentration in samples can be determined by extrapolation to this standard curve.

## ASSAY CONDITIONS

Plasma, serum, culture media, tissue homogenate, CSF, urine or any biological fluid can be assayed as long as the level of the sample is high enough the for the sensitivity of the kit to detect it.

**CAUTION:** Phoenix Pharmaceuticals guarantees that its products conform to the information contained in this publication. The purchaser must determine the suitability of the product for its particular use and establish optimum sample concentrations.

## Assay Principle



## LIST OF COMPONENTS

*Store all components at 4°C. DO NOT FREEZE.*

1. **20x Assay Buffer concentrate (50ml)**.....**Catalog No. EK-BUF**
2. **96 Well Anti-Human C-Reactive.....Catalog No. EK-Plate-072-62**  
Protein Capture Antibody-Coated Plate (*1 plate*)
3. **Human C-Reactive Protein Standard.....Catalog No. EK-S-072-62**  
(*500ng/vial*) (*100µl*)
4. **Anti-Human C-Reactive.....Catalog No. EK-D-072-62**  
Protein Detection Antibody (*1 vial*)
5. **Human C-Reactive Protein.....Catalog No. EK-PC-072-62**  
Positive Control (*2 vials*)
6. **Substrate Solution (TMB) (12ml)**.....**Catalog No. EK-TMB**
7. **Stop Solution 2N HCl (15ml)**.....**Catalog No. EK-HCl**
8. **Acetate plate sealer (APS) (3 pieces)**.....**Catalog No. EK-APS**
9. **Assay Diagram (1 sheet)**

## MATERIALS REQUIRED BUT NOT SUPPLIED

- Micropipettor(s) and disposable pipette tips
- Multi-channel pipette capable of dispensing 50-100µl
- Solution Reservoir (*recommended*)
- Microtiter plate washer (*recommended*)
- Orbital plate shaker capable of 300-500 rpm (*recommended*)
- Microtiter plate reader capable of absorbance measurement 450nm
- Well-closed containers (15ml tubes or more in capacity)
- Absorbent material for blotting

## REAGENT PREPARATION

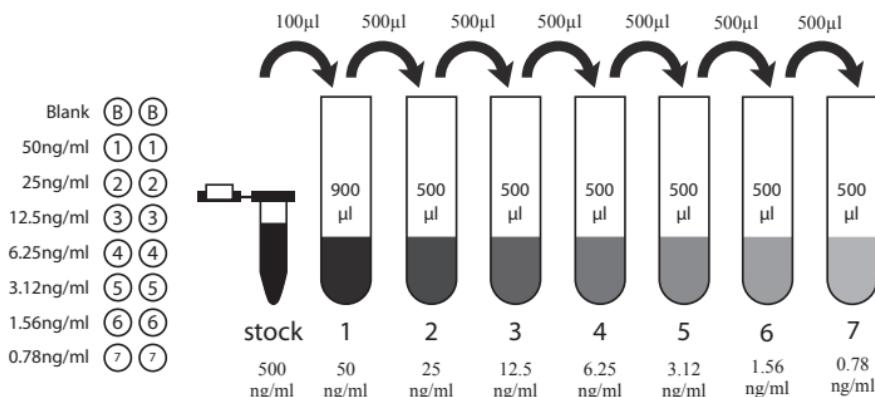
Note: *The kit should be equilibrated to room temperature (20-23°C) before opening any vials and starting the assay. It is highly recommended that the solutions be used as soon as possible after rehydration.*

1. **1x Asssay buffer:** Dilute the **20x** assay buffer concentrate with 950ml of distilled water. This assay buffer will be used to wash the plate and reconstitute all of the other components in this kit. If crystals are observed in the **20x** Assay buffer, warm the bottle in a 37°C water bath for approximately, 30 minutes or until the crystals disappear. After preparation, store **1x** assay buffer at 4°C.
2. **Anti-Human C-Reactive Protein Detection Antibody:** Rehydrate anti-Human C-Reactive Protein Detection Antibody with 100µl of **1x** assay buffer, vortex (centrifuge the tube to dislodge powder from the cap or walls). Dilute biotinylated anti-Human C-Reactive Protein Detection Antibody to 1:450 and mix thoroughly before use.
3. **Human C-Reaction Protein Positive Control:** For Human C-Reactive Protein Positive Control, add 200µl of **1x** assay buffer (centrifuge the tube). Vortex thoroughly.

## HUMAN C-REACTIVE PROTEIN STANDARD PREPARATION

1. For recombinant Human C-Reactive Protein standard, add 900µl **1x** asssay buffer, vortex. Allow the solution to sit at least 10 minutes at room temperature (20-23°C) to completely dissolve in solution. Vortex and centrifuge before use. The concentration of this stock solution is 500ng/ml.
2. Prepare Human C-Reactive Protein standard solutions as follows:

Standard No.	Std. volume	Assay Buffer	Concentrations
Stock	100µl	900µl	500ng/ml
#1	100µl of Stock	900µl	50ng/ml
#2	500µl of #1	500µl	25ng/ml
#3	500µl of #2	500µl	12.5ng/ml
#4	500µl of #3	500µl	6.25ng/ml
#5	500µl of #4	500µl	3.12ng/ml
#6	500µl of #5	500µl	1.56ng/ml
#7	500µl of #6	500µl	0.78ng/ml



## HUMAN C-REACTIVE PROTEIN ELISA PROTOCOL

1. Thoroughly read this protocol before performing an assay. Allow all reagents to come to room temperature (20-23°C) prior to the start of the assay.
2. Remove Capture Antibody-Coated Plate from its zip-lock foil pouch. Remove unneeded strips from the plate frame, reseal them in the foil pouch, and return the foil pouch to 4°C.
3. Wash each well with 300µl of **1x** assay buffer. Allow it to sit for at least five minutes. Discard the buffer, invert and blot dry plate. Do not let wells dry before proceeding to the next step.
4. Leave wells A-1 and A-2 empty as **Blank**.

## C-REACTIVE PROTEIN (HUMAN) ELISA KIT PROTOCOL

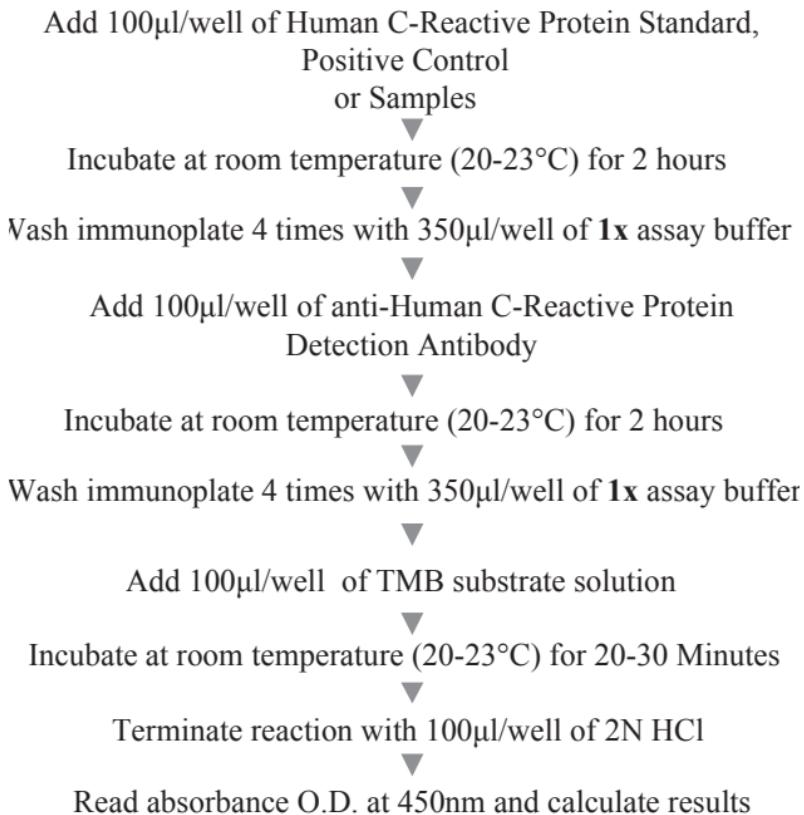
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5. Add 100 $\mu$ l of the prepared Human C-Reactive Protein Standard solutions from #7 to #1 (reverse order of serial dilution) in duplicate to each well.
6. Add 100 $\mu$ l of Human C-Reactive Protein positive control solution in duplicate.
7. Add 100 $\mu$ l of samples (diluted/undiluted) in duplicate into their designated wells.
8. Seal the immunoplate with acetate plate sealer (APS). Incubate for 2 hours at room temperature (20-23°C) on a plate shaker (300-400 rpm).
9. Before washing the plate, remove the plate sealer carefully. Completely discard the liquid from wells. Wash each well with 300-350 $\mu$ l assay buffer four times. At the end of the wash, discard the buffer, invert the plate, and tap on a clean absorbent towel.
10. Add 100 $\mu$ l anti-Human C-Reactive Protein Detection Antibody into each well. Reseal the immunoplate with plate sealer and incubate for 2 hours at room temperature (20-23°C) on a plate shaker (300-400rpm).
11. Wash 4 times with the 1x assay buffer as described in step 9.
12. Add 100 $\mu$ l substrate solution (TMB) provided in this kit into each well. Reseal the plate with plate sealer to protect from light and incubate the plate for 20-30 minutes at room temperature (20-23°C) on a plate shaker (300-400 rpm).
13. Add 100 $\mu$ l Stop Solution (2N Hydrochloric Acid) into each well to stop the reaction. The color in the well should change from blue to yellow. If the color change does not appear to be uniform, gently tap the plate to ensure thorough mixing. Proceed to the next step within 20 minutes.
14. Read Absorbance O.D. at 450nm using a Microtiter Plate Reader.

**ADDITIONAL RECOMMENDED PROCEDURAL NOTES:**

- Reagents of different lot numbers should not be mixed.
- Recheck the reagent labels when loading the plate to ensure that everything is added correctly.
- Unused microplate strips should be placed in the foil pouch with a dessicant and stored at 4°C. Do not allow moisture to enter the wells.
- When handling the plate, avoid touching the bottom.
- Manual washing may cause high duplicate coefficient variations. To reduce this factor, liquid from the plate should be removed by inverting and blotting the plate on an absorbent material.
- If the room temperature is not within the suggested range (20-23 °C), variations in results may occur.
- The same reservoir for the reagents may be reused if the reservoir is washed well with distilled water before each use.
- Each laboratory must determine the appropriate dilution factors for the samples to be measured to ensure that the samples are within the dynamic range of the standard curve.
- High levels of interfering proteins may cause variations within the sample results, therefore, it is imperative to select the appropriate sample preparation procedure to obtain optimal results.
- Each time a new tip is used, make sure the tip is secure and free of air bubbles. For better intra-assay variation, aspirate and expel a reagent or sample back into the container a few times prior to loading.
- Avoid submerging the whole tip into reagents because droplets can accumulate at the end of the tip causing an excess of reagent to be loaded into the well. This can lead to poor results.
- For optimal results, an orbital plate shaker capable of 300-500 rpm is recommended for all incubations.
- Modification of the existing protocol (i.e. standard dilutions, pipetting technique, washing technique, incubation time or temperature, storage conditions, and kit expiration) may affect the sensitivity and specificity of the test.

## SUMMARY OF ASSAY PROTOCOL



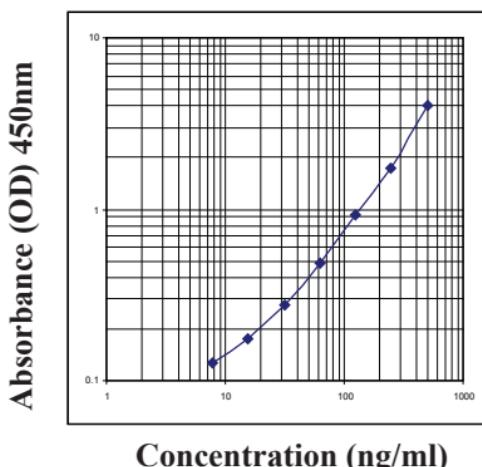
## CALCULATION OF RESULTS

Plot the standard curve on log-log graph paper. Known concentration of Human C-Reactive Protein Standard and its corresponding O.D. reading is plotted on the log scale (X-axis) and the log scale (Y-axis), respectively. The standard curve shows a correlated relationship between Human C-Reactive Protein concentrations and the corresponding O.D. absorbance. As the standard concentration increases, the intensity of the yellow color, and in turn the O.D. absorbance, increases.

The concentration of Human C-Reactive Protein in a sample is determined by plotting the sample's O.D. on the Y-axis, then drawing a horizontal line to intersect with the standard curve. A vertical line dropped from this point will intersect the X-axis at a coordinate corresponding to the Human C-Reactive Protein concentration in the unknown sample.

Refer to QC Data sheet for acceptable values of the positive control.

**Human C-Reactive Protein Standard Curve**



## STORAGE

1. Store the kit at 4°C upon receipt. The kit should be equilibrated to room temperature (20-23°C) before assay.
2. Store **1x** assay buffer at 4°C.
3. Remove any unneeded strips from Human C-Reactive Protein Antibody-Coated plate, re-seal them in zip-lock foil and keep at 4°C.
4. Keep rehydrated solution of Human C-Reactive Protein Standard, anti-Human C-Reactive Protein-HRP Detection Anti-body at 4°C. Prepare only the required amount.

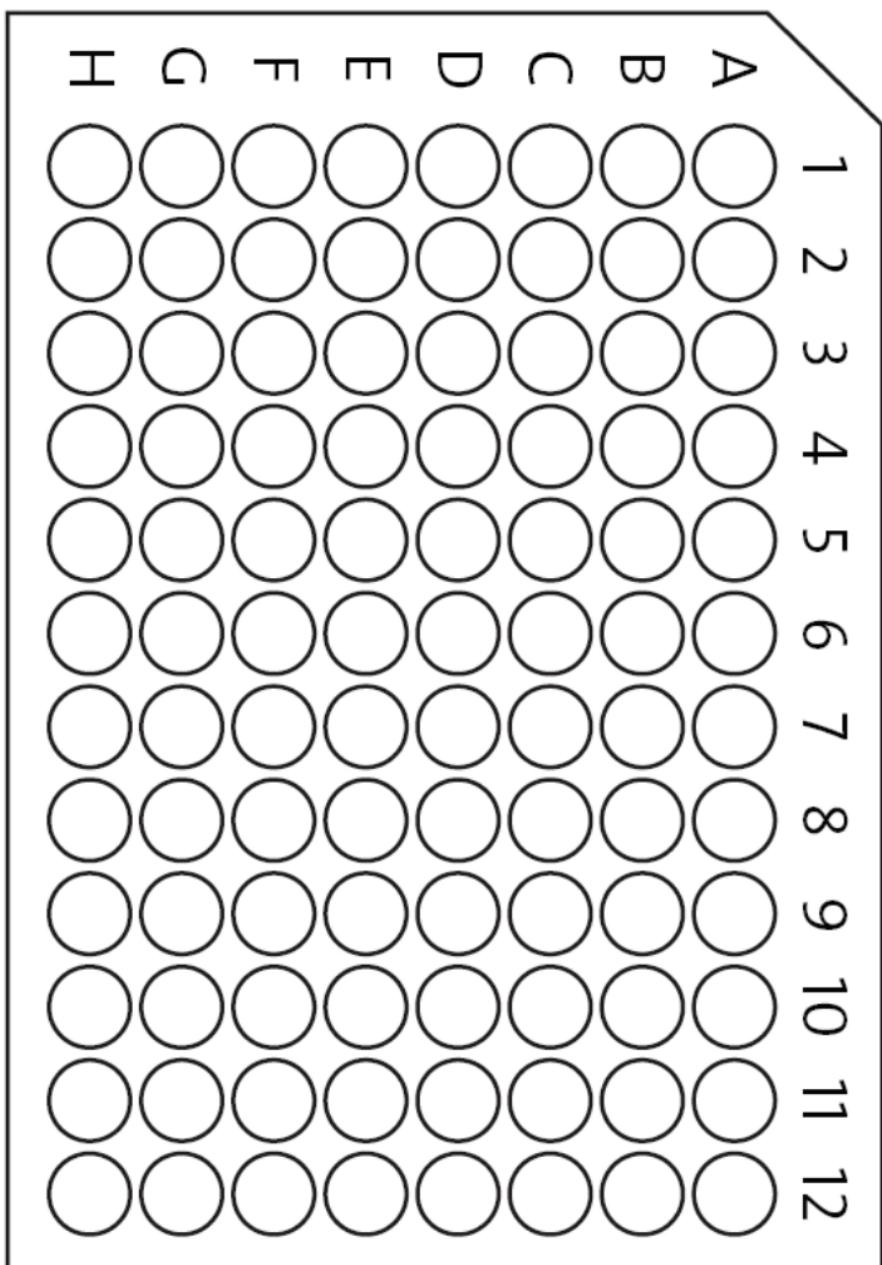
## NOTE:

1. It is recommended that the solutions be used on the same day of rehydration.
2. Unextracted serum samples of normal subjects are to be diluted with **1x** assay buffer.
3. After adding Stop Solution, read the plate within 20 minutes.

## REFERENCES

1. Vlankis JE. Human C-reactive protein: expression, structure, and function. Molecular Immunology. 2001 Aug; 38(2-3):189-97.
2. Sursh MV, Singh SK, Fergusson, DA Jr., Agrrawal A. Role of the property of C-reactive protein to activate the classical pathway of complement in protecting mice from pneumococal infection. Journal of Immunology. 2006 April 1:176(7):4.69-74.
3. Protective effect of ghrelin on left ventricular remodeling in spontaneously hypertensive rats is associated with the peroxi some proliferator activated-receptor gamma dependant pathway. LI Zhao, ZHU Xiao-ying, LI Meng, BAI Ying-long, HU Jian LI Zhao.Chinese Medical Journal, 2008, Vol. 121 No. 22 : 2299-2304.

**ASSAY DIAGRAM**



**NOTES**